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N-STORM with DNA-PAINT for Reliable Multicolor & 3D Single Molecule Localization Imaging

New DNA-PAINT immunolabeling reagents available from Ultivue Inc. enables photobleach-free single molecule localization imaging using the Nikon N-STORM 4.0 super-resolution microscope. Multi-color and 3D imaging with a depth of more than 5 μm is demonstrated here.

What is DNA-PAINT?

The past decade has seen the introduction of super-resolution STORM¹ and similar single molecule localization microscopies for nanoscale imaging. These techniques generally rely upon the photochemical manipulation of fluorophores to induce long-lived non-fluorescent 'dark' states, allowing the emission profiles of non-overlapping single emitters to be identified. In contrast, Point Accumulation for Imaging in Nanoscale Topography (PAINT) is a set of techniques that utilizes short-lived reversible binding of fluorophores to localize single molecules². The advantage of this approach is that it virtually negates the effects of photobleaching as fatigued fluorophores are continuously replaced, allowing each molecule of interest to be sampled a theoretically unlimited number of times.

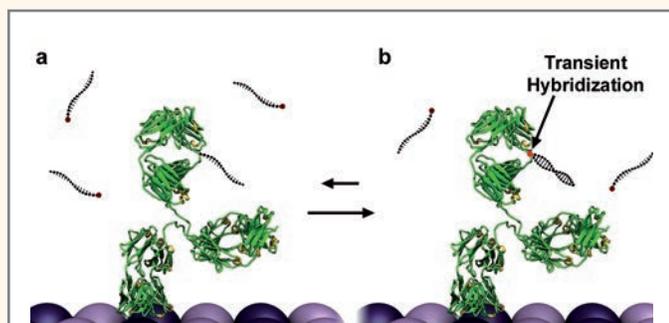


Figure 1. Illustration of the DNA-PAINT Concept. Imager strands reversibly bind to docking strands on the secondary antibody with short on-times. Although fluorescence is excited from imager strands free in solution, immobilization of a fluorophore allows for a much greater amount of signal to be integrated when average binding on-time is synced to camera exposure, usually about 25-40 milliseconds per frame.

DNA-PAINT³ is a variation of PAINT utilizing conventional primary-secondary antibody immunolabeling techniques. However, instead of a fluorophore, the secondary antibody is conjugated to short single-stranded DNA oligomers known as **docking strands**. The imaging buffer contains complementary oligomers, known as **imager strands**, labeled with fluorophore. Imager strands demonstrate binding on-times in the tens of milliseconds, during which time the observed fluorescence is greatly enhanced over the background as the fluorophore becomes temporarily immobilized (Figure 1), ideal for single molecule localization.

Complete DNA-PAINT reagent kits, including secondary antibodies, imager strands, and buffers, are commercially available from Ultivue Inc. (www.ultivue.com), providing a convenient and consistent solution.



N-STORM 4.0 for PAINT Imaging

DNA-PAINT can be performed on the Nikon N-STORM 4.0 without alteration to the microscope, requiring only minor changes to standard immunolabeling protocols. DNA-PAINT kits are provided with both goat anti-mouse and goat anti-rabbit secondary antibodies, each with a unique docking strand sequence. This allows for the application of imager strands labeled with different fluorophores for simultaneous imaging, or with the same fluorophore for sequential imaging (Exchange-PAINT³). Note that using different imager strands with the same fluorophore requires washing one set out before labeling with the other.

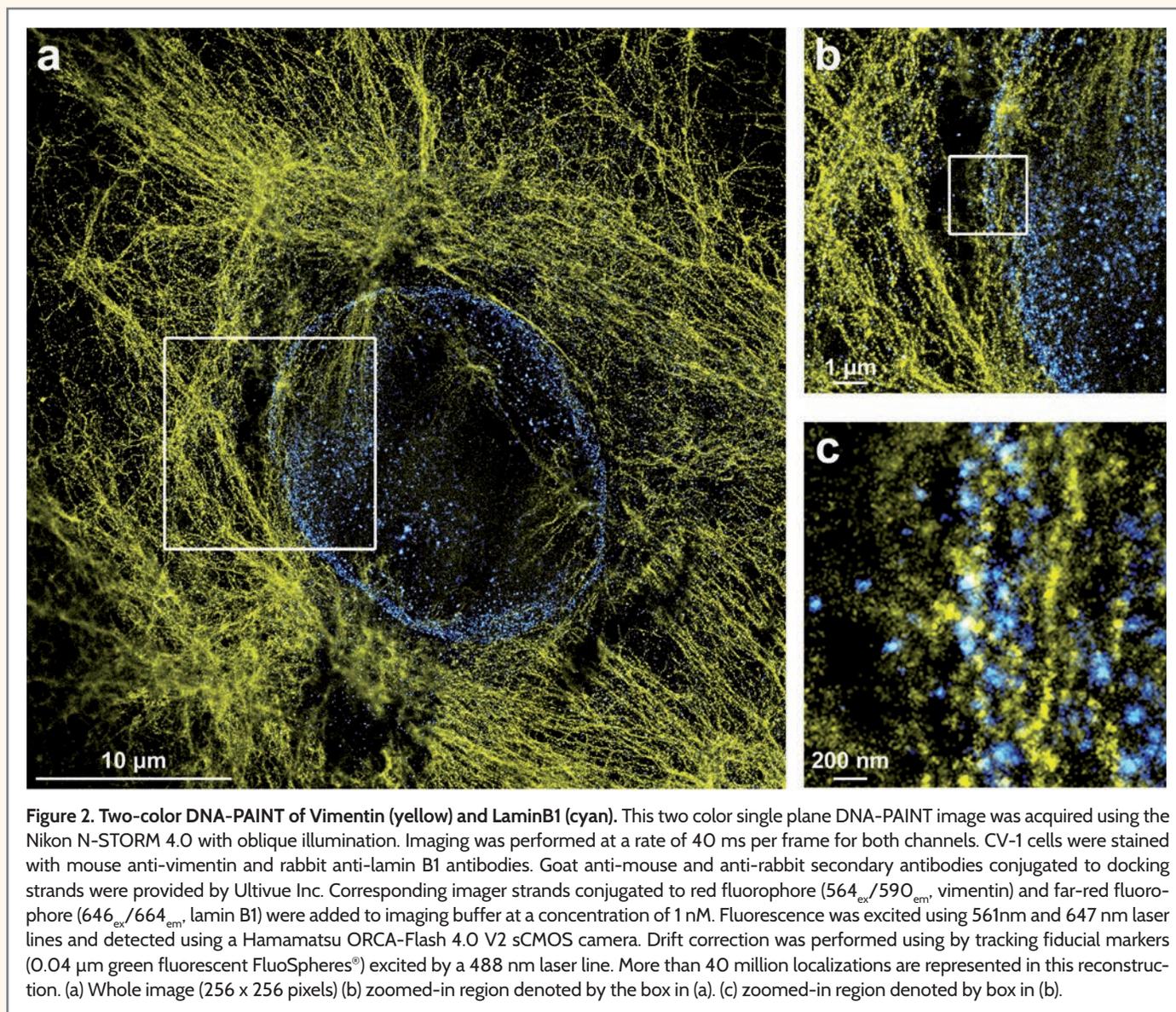
Multi-Color DNA-PAINT

Here we demonstrate wash-free simultaneous two-color DNA-PAINT imaging with spectrally distinct fluorophores. Up to three color channels are available on the N-STORM 4.0, allowing for imaging up to three colors, or imaging of two

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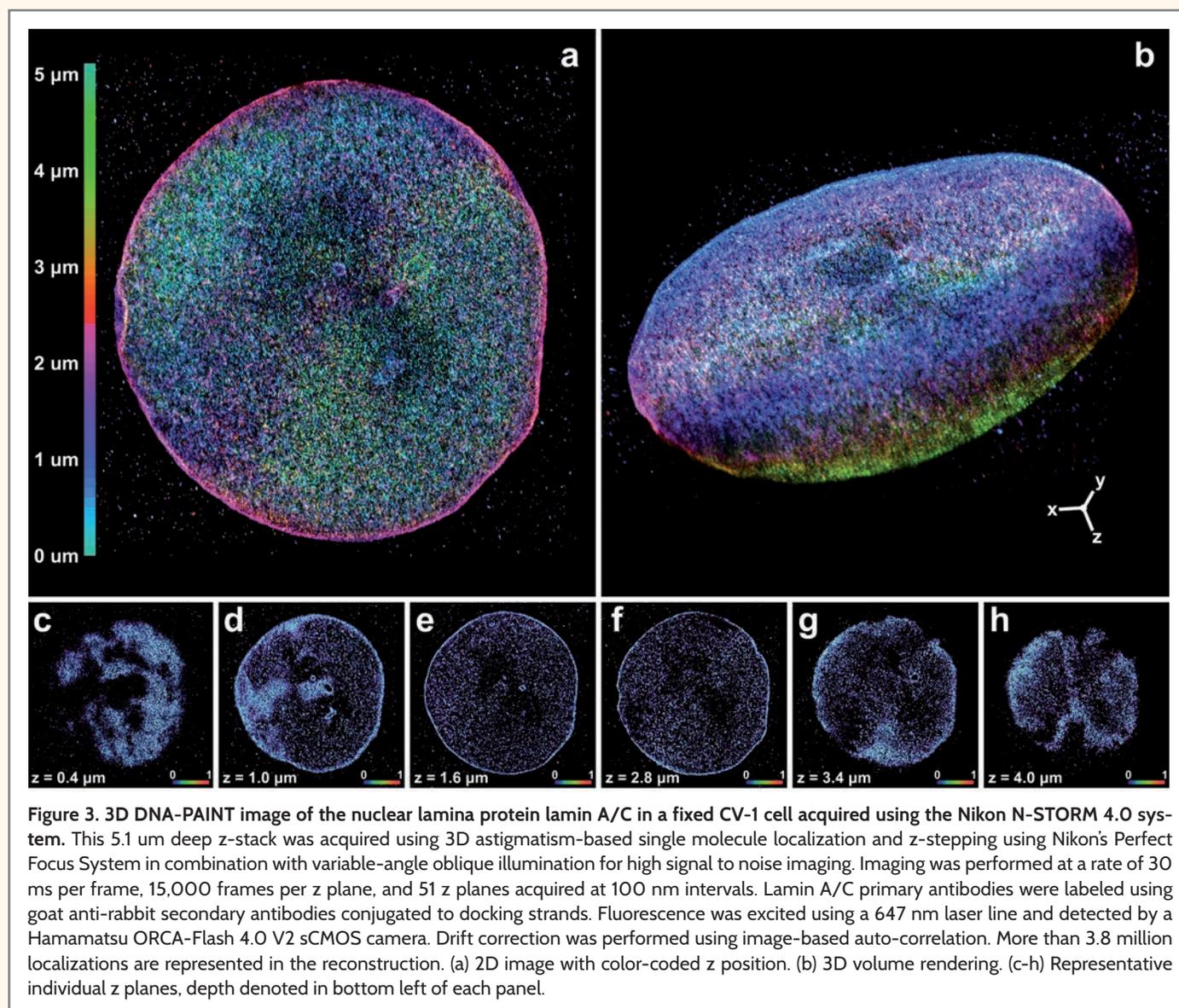
colors with the third color channel for tracking fiducial markers for drift correction, as in Figure 2.

Multi-color DNA-PAINT has several inherent advantages over multi-color dSTORM. DNA-PAINT probes exhibit similar brightness in all tested color channels – providing consistent resolution performance in each channel and competitive with Alexa Fluor 647, the current standard in the field. Multi-color dSTORM typically relies upon fluorophores with markedly reduced performance for imaging in the green (e.g. Atto 488) and red (e.g. Cy3B) channels. Localization precision when using DNA-PAINT is dependent on the binding kinetics of the imager strand and the brightness of the fluorescent probe, not requiring specialized and short-lived (1-2 hours) reducing/oxidizing buffers. Imager strands are simply diluted in the provided salt

buffer – with the resultant imaging buffer good for up to several days at room temperature and with very little drop-off in the number of molecules localized in each frame during that time.

3D DNA-PAINT

The N-STORM 4.0 provides a proprietary z-stepping mechanism for single molecule imaging using Nikon's Perfect Focus System. Single molecule z-stacks 5+ μm thick can be acquired with high quality. Enhanced signal to noise and background suppression is provided through a specialized combination of TIRF and oblique illumination, where the angle of illumination changes with imaging depth, allowing for more selective excitation of the z plane of interest. Drift correction for the data in Figure 3 was performed using image-based auto-correlation analysis.



While z-stacking can be performed using conventional STORM labels and buffers on the N-STORM 4.0, the DNA-PAINT approach provides improved performance as the user no longer has to balance photobleaching (occurring throughout the sample volume) with the acquisition period for each z position.

Outlook

DNA-PAINT probes from Ultivue Inc. provide a convenient and standardized method for performing high quality 3D and multicolor single molecule imaging on Nikon's N-STORM 4.0 system, and with conventional Immunolabeling techniques. Combination of DNA-PAINT probes with other dyes validated for PAINT imaging allows for potential three-color single mol-

ecule imaging, and in 3D. We also expect the future development of a wider variety of DNA-PAINT probes to continue to expand the number of potential applications for this technique.

References

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2. Sharonov, A. and Hochstrasser, R.M. Wide-field subdiffraction imaging by accumulated binding of diffusing probes. *Proc. Natl. Acad. Sci. U.S.A.* 103, 18911-18916 (2006).
3. Jungmann, R., Avendano, M.S., Woehrstein, J.B., et al. Multiplexed 3D cellular super-resolution imaging with DNA-PAINT and Exchange-PAINT. *Nature Methods* 11, 313-318 (2014).