Shedding New Light On MICROSCOPY



APPLICATION NOTE

ECLIPSE Ji Digital Inverted Microscope AX Confocal Microscope System General Analysis 3 Image analysis Software module

3D image analysis reveals drug-induced cellular senescence and increased nucleolus volume: Efficient imaging methods to maximize data acquisition

High content analysis incorporates multiple measurements such as cell morphology and fluorescence intensity of specificallylabelled cellular features to assess biological activity in response to set of conditions, and is a powerful approach for applications like drug compound screening. Wide-field microscopy offers relatively high imaging throughput, which is ideal for screening drug candidate compounds, while confocal laser scanning microscopy enables three-dimensional visualization of structures and volumetric quantification to obtain deeper insight into compound responses. Using both wide-field and confocal laser scanning microscopy, drug discovery and life science research can be advanced more efficiently. In this application note, we introduce an imaging method that efficiently utilizes both 2D and 3D analysis of wide-field and confocal images to reveal how drug-induced cellular senescence impacts the nucleolus.

Keywords: cellular senescence, high content analysis, wide-field microscopy, confocal microscopy, 3D analysis, volume measurement, nucleolus, DNA damage, yH2AX, granule analysis, drug discovery

Sample preparation and experimental results

2D analysis (Fig. 1): A549 cells were treated with Doxorubicin (DOX) at concentrations of 50, 100, 200, and 400 nM to induce senescence. The senescent A549 cells were seeded in a 96-well plate, and nuclei, nucleoli, and γH2AX granules were fluorescently stained. First, we identified compound concentrations that showed changes by 2D analysis of nucleoli and γH2AX using a Wide-field microscope. **3D** analysis (Fig. 2): Next, we used a confocal microscope to acquire images of DOX concentrations (0, 100, 400 nM) and ran 3D analysis of nucleoli and nucleoli. 2D analysis showed that the area of the nucleoli were similar at different DOX concentrations (Fig.1 (E)). However, 3D analysis revealed that the volume of the nucleoli increased at higher compound concentrations (Fig. 2(E)). This clearly indicates that 3D analysis of nucleoli volume is a critical analytical parameter in the study of cellular senescence.



Fig. 1: Wide-field images, 2D binarized images, and analysis results Objective: PLAN APO λ D 20x (NA0.8), (A, B, C) Images of A549 cells, blue: nuclei (DAPI), yellow: nucleoli (Nucleolus Bright Red), red: γ H2AX. Middle row: Enlarged images of representative regions. Lower row: Mask images (cyan: nucleus mask, yellow: nucleolus mask, red: γ H2AX mask). Scale bar: 100 µm (upper row), 20 µm (middle and lower rows). (D-J) 2D analysis results. (X-axis: DOX concentration (nM), Y-axis: population statistics). (D, E) The area of the nuclei and nucleoli increased approximately two-fold with Doxorubicin (DOX) treatment, but showed no significant difference above 50 nM DOX. (F) On the other hand, the number of nucleoli was maximized at 100 nM DOX, and (G) the fluorescence intensity of Nucleolus Bright Red within the nucleus mask showed a concentration-dependent increase. (H, I, J) The analysis results of γ H2AX all showed the maximum values at 200 nM DOX.

Fig. 2: Confocal 3D images, 3D binarized images, and analysis results Objective: CFI Apo LWD Lambda S 40XC WI (N.A. 1.15), Scan mode: Galvano scanner, Resolution: 1024 x 1024 pixel
(A) 3D image of the nucleus and nucleolus of A549 cells treated with 400 nM DOX (color-coded by Z depth: Z=0-5 µm (blue), Z=5-15 µm (green), Z=15-20 µm (orange)). (B) 3D binarized image (XY plane), (C) XZ plane image of (B), (B, C) magenta: nucleus mask, blue: nucleolus mask.). Scale ruler: 10 µm. Nucleoli exhibit diverse morphologies, and 3D analysis revealed that DOX treatment increased nucleolus volume in a concentration-dependent manner (E).

Fast imaging with a large field of view and high resolution to capture tiny granules of yH2AX

ECLIPSE Ji built-in camera and 20x lambda D objective combine to achieve a resolution per pixel of $0.316 \ \mu m/pixel$

Even small granular structures of less than 1 μ m in size can be resolved with a low-magnification 20x objective, allowing the capture of a large number of cells in a single shot with the potential for quantitative statistical analysis.



2D & 3D high-content analysis to maximize data acquisition efficiency





Wide-field: Study populations

- ✓ Drug Screening
- ✓ Drug concentration considerations



Confocal: Explore details

- ✓ Gain subcellular insight
- ✓ Make new discoveries
- ✓ Conduct 3D analysis

Imaging Modality	Nucleus	Nucleolus	γΗ2ΑΧ	Features
Wide-field	Area	Fluorescence	Number	✓ Population
(2D)		Intensity	• Area	overview
			• Fluorescenc	✓ Fast image
			e Intensity	acquisition
Confocal	Volume	Volume		✓ 3D image
(3D)				✓ Volume info

Wide-field

Wide-field

Confocal

Table. 1: Analytical parameters and optimal imaging methodfeatures for studying cellular senescence





- ✓ Rapidly capture images
- Low-magnification objective captures many cells for quantitative statistical analysis
- ✓ Good for drug screening
- \checkmark Lighter data volume and easy analysis





Confocal: Details

- ✓ Three-dimensional visualization
- ✓ 3D analysis
- ✓ Volume information



3D: 40x Water immersion objective

3D analysis by constructing 3D images from Z-stack images



[DOX]: 0, 100, 400 nM Experimental groups: 3 conditions 3 images/well, 3 wells Total: 9 images

Detection region	Fluorescence label	Ex/Em (nm)			
Nucleus	DAPI	345/455			
Nucleolus	Nucleolus Bright Red	537/605			
γH2AX (DNA damage)	DNA Damage Detection Kit - γH2AX - Deep Red	646/668			
Light source and detector					
Epi-fluorescence (D-LEDI2), Ji Built-in camera (FOV 25mm)					
Objective (N.A.)					
CFI Plan Apochromat Lambda D 20X (N.A. 0.8)					
Magnification	Field of view (FOV), R	esolution			
20X	0.88 x 0.88 mm / image 2800 x 2800 pixel				
# of points	Z-stacks				
2 points/well	1.825 µm x 3 steps (Range:3.75 µm)				
# of wells	Drug (Doxorubicin) concentrations				
15 wells	0, 50, 100, 200, 400 nM				
# of images	Image processing				
30 images	Constructing EDF images from Z-stack images				

Detection region	Fluorescence label	Ex/Em (nm)			
Nucleus	DAPI	345/455			
Nucleolus	Nucleolus Bright Red	537/605			
Light source and detector					
Confocal Laser (AX), DUX-VB 4Ch Detector Unit (FOV 25mm)					
Objective (N.A.)					
CFI Apo LWD Lambda S 40XC WI (N.A. 1.15)					
Magnification	Field of view (FOV), Resol	ution, Others			
	0.44 x 0.44 mm / image				
40X	1024 x 1024 pixel				
	Galvano, Averaging: 2				
# of points Z-stacks					
3 points/well 0.32 µm x 45-70 steps (Range:14 – 22		ge:14 – 22 µm)			
# of wells	Drug (Doxorubicin) concentrations				
3 wells	0, 100, 400 nM				
# of images	Image processing				
9 images	Constructing a 3D image from Z-stack images				

Table. 2: Detection area, fluorescent labels, and conditions for image acquisition using a wide-field microscope for Fig 1. Table. 3: Detection area, fluorescent labels, and conditions for image acquisition using a confocal laser-scanning microscope for Fig 2.

Fast, accurate imaging for new discoveries

In this experiment, we used a wide-field microscope to capture a large number of images in a short time, allowing us to study the effects of a drug on variably-treated populations across the entire plate. A single, fully in-focus EDF image was constructed from three Z-stack images, and the nuclei, nucleoli, and yH2AX granules were binarized and measured. γH2AX granules appeared with maximum count, area, and fluorescence intensity at 200 nM DOX treatment. On the other hand, the number of nucleoli and their fluorescence intensities were maximal with 100 and 400 nM Dox treatment, respectively. The area of nucleoli showed no difference with drug concentration above 50 nM. To then study subcellular details, confocal images of A549 cells at three different drug concentrations were acquired. The 3D confocal images were binarized and the volumes of the nuclei and nucleoli were measured. The results of the 3D analysis revealed that high concentrations of Doxorubicin increased nucleolus volume.

Cell staining protocol

DNA Damage Detection Kit - γH2AX - Deep Red

* This is slightly different from the method described in the G267 instruction manual.

- 1) Remove the cell culture supernatant, add 4% PFA solution (100 µl/well) to each well, and incubate at room temperature for 3 minutes.
- 2) Remove the PFA solution and wash the cells three times with PBS (100 μ l/well).
- 3) add 0.1% Triton X-100/PBS solution (100 µl/well) and incubate at room temperature for 30 minutes.
- 4) Remove the supernatant and wash the cells twice with PBS $(100 \ \mu l/well)$.
- 5) Add Blocking Solution (100 µl/well) and incubate for 20 minutes at room temperature.
- 6) Remove the supernatant and wash the cells twice with PBS $(100 \ \mu l/well)$.
- 7) Add γ H2AX staining solution (100 μ l/well) and incubate overnight at 4 degrees.
- 8) Remove the supernatant and wash the cells twice with PBS $(100 \ \mu l/well)$. 9) Add Secondary antibody staining solution (100µl/well) and incubate for 1 hour at room temperature. 10)Remove the supernatant and wash the cells twice with PBS $(100 \ \mu l/well)$.

Summary

- \checkmark Fast, 2D statistical analysis with wide-field microscopy
- ✓ Three-dimensional structural examination and 3D analysis with confocal laser scanning microscopy
- ✓ Volumetric information leads to new discoveries
- ✓ Efficient imaging was combined 2D and 3D analysis
- \checkmark Nucleolus volume is a useful analytical parameter in the study of cellular senescence
- Combined imaging modalities allow accurate detection of drug effects
- ✓ Seamless 2D & 3D analysis from drug discovery to pharmacological effects with a single ECLIPSE Ji

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Sample Preparation Protocol

Please refer to the Nikon ECLIPSE Ji Application Note for the A549 cell senescence induction and sample preparation protocol "Labelfree quantitative analysis of cellular senescence and high-content

Nucleolus Bright Red

- 1) After γ H2AX staining, remove the supernatant and add a mixture of adjusted Nucleolus Bright working solution and DAPI (100 μ /well) and incubate at room temperature for 5 minutes. * Please refer to the instruction manual of N512. Nucleolus Bright Red: 1000x dilution, DAPI: 1000x dilution https://www.dojindo.co.jp/manual/N511 N512/
- 2) The supernatant is removed and the cells are washed twice with PBS (100 μ l/well).

Reagents, Product name	Product number	Supplier
-Cellstain®- DAPI solution	D523	Dojindo Laboratories
DNA Damage Detection Kit - γH2AX - Deep Red	G267	Dojindo Laboratories
Nucleolus Bright Red	N512	Dojindo Laboratories

References

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imaging of granules"

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Product information

ECLIPSE Ji Digital Inverted Microscope with AX Confocal Microscope System

The ECLIPSE Ji is capable of fast, high-resolution image acquisition using its built-in camera and epi-fluorescence light source. This digital inverted microscope is optimized for 2D high content analysis, with the ability to upgrade to confocal when required. From drug screening using fast wide-field imaging to understanding three-dimensional structures and 3D analysis using confocal images, you can obtain seamless results with this single unit.





General Analysis 3 image analysis software module

By simply combining analysis blocks, nucleus and nucleolus regions can be easily binarized and measured, allowing flexible image analysis.

The "ConnectCells ^{3D}" function allows you to easily perform 3D analysis in steps similar to a 2D analysis.



