

## Protein components of virus particles in SARS-CoV-2-infected cells imaged by N-STORM super-resolution microscope

Elucidating the mechanism of SARS-CoV-2 proliferation is important for the development of prevention and treatment strategies for COVID-19. In cells infected with SARS-CoV-2, the expression levels, ratios, and intracellular localizations of each virus-constituting protein change as virus particles are formed. Although endosomes with numerous intraluminal vesicles (multivesicular bodies; MVBs) and lysosomes are now being recognized as locations where virus particles are assembled, it is unclear how each viral protein accumulates within them.

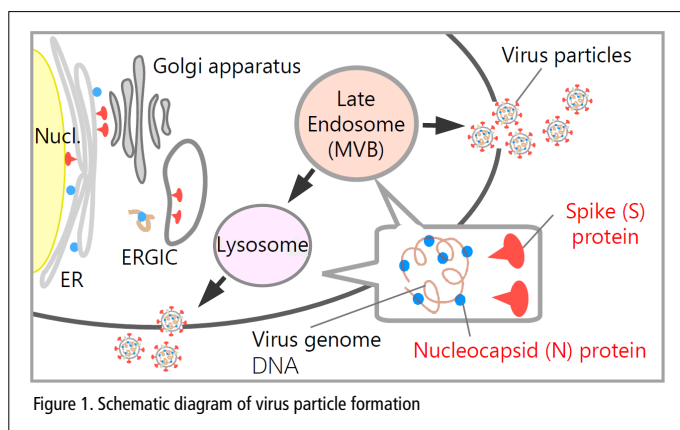
Ms. Yoko Ishida, Dr. Shinichiro Okamoto, Dr. Megumu Takahashi, and Prof. Hiroyuki Hioki (Department of Neuroanatomy, Graduate School of Medicine, Juntendo University) observed the localization of the spike (S) protein and nucleocapsid (N) protein within MVBs and lysosomes using both a confocal microscope and N-STORM super-resolution microscope.

In this application note, we present images of virus particle protein components in SARS-CoV-2-infected cells successfully captured by N-STORM.

Keywords: SARS-CoV-2, COVID-19, spike (S) protein, nucleocapsid (N) protein, N-STORM super-resolution microscope

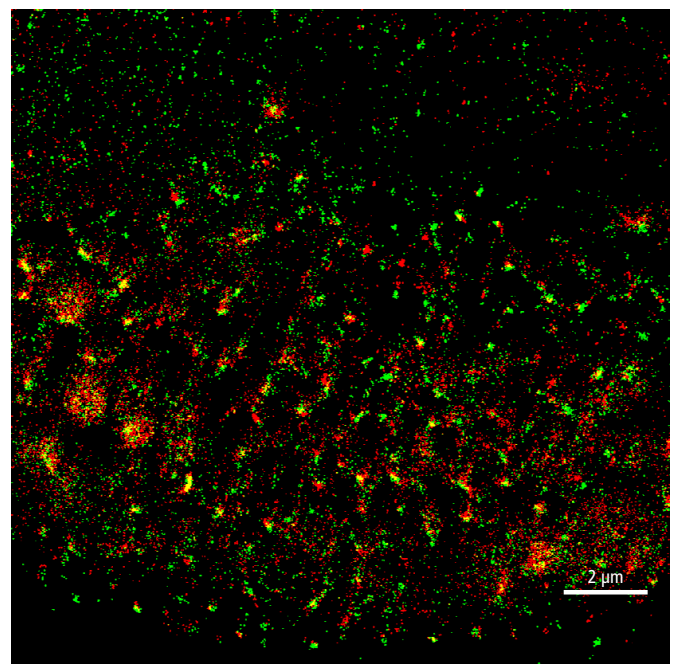
### Overview

The process of virus particle formation within the MVBs and lysosomes of SARS-CoV-2-infected cells has been extensively analyzed using electron microscopy. While electron microscopy has the advantage of observing various structures based on electron density, immunoelectron microscopy or correlative light-electron microscopy are required to analyze the localization of a specific protein. Super-resolution microscopy combined with fluorescence immunostaining enables localization of multiple proteins with high spatial resolution. In this experiment, we observed the localization of S and N proteins inside MVBs with a resolution of tens of nanometers using N-STORM super-resolution microscopy (Fig. 1).

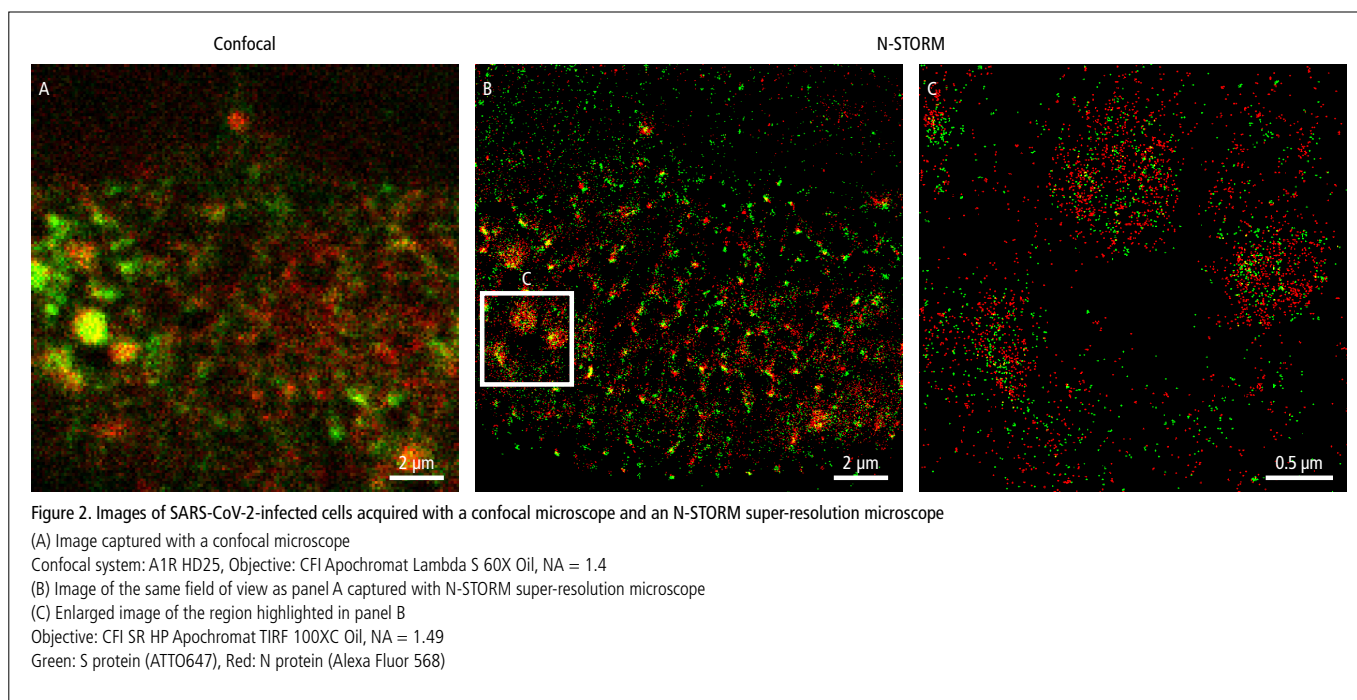


### Materials and Methods

Vero cells were infected with SARS-CoV-2 and fixed with 4% paraformaldehyde 24 hours post-infection. Subsequently, multiple immunofluorescence staining was performed with anti-S protein VHH antibody, anti-N protein VHH antibody, and anti-TSG101 IgG antibody (MVB marker). Following the observation of TSG101-positive regions using confocal microscopy, the same regions were examined using an N-STORM super-resolution microscope (Fig. 2).



An image of SARS-CoV-2-infected cells acquired with an N-STORM microscope



## Results

Observation of the infected cells with a confocal microscope revealed that most of the fluorescence signals for S and N proteins were colocalized (Fig. 2A). Observation of the same field using an N-STORM super-resolution microscope elucidated the intracellular localization of S and N proteins at higher spatial resolution (Fig. 2B). In the enlarged image, the fluorescence signals generated by the immunolabeled S and N proteins appear as separate but proximally-located bright spots (Fig. 2C).

## Summary

Super-resolution microscopy is a technique that enables observation of detailed structures beyond the diffraction limit of light, and it achieves optical observation of ultra-fine morphologies that could previously only be observed with electron microscopy. Furthermore, N-STORM makes it possible to perform super-resolution localization of multiple antigens within the same sample. In this study, we observed the intracellular distribution of S and N proteins within SARS-CoV-2-infected cells at high spatial resolution. We have demonstrated the capability of N-STORM toward analyzing the intracellular localization of virus particle protein components with high precision, thus contributing to the elucidation of the virus particle formation mechanism.

## Acknowledgments

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## References

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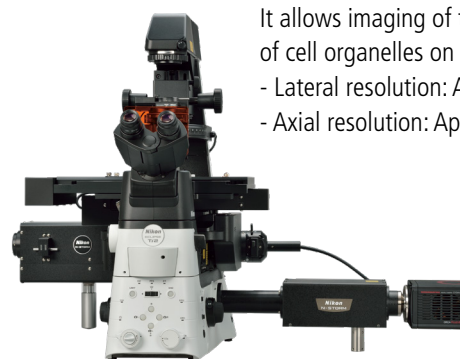
## Product information

### N-STORM Super Resolution Microscope

The N-STORM utilizes a localization technique called STochastic Optical Reconstruction Microscopy (STORM) to achieve a resolution of ten times that of conventional light microscopes.

It allows imaging of the structure of cell organelles on a molecular level.

- Lateral resolution: Approx. 20 nm
- Axial resolution: Approx. 50 nm



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